

## Data-driven models of sequence landscapes

### Martin Weigt

#### Laboratoire de Biologie Computationnelle et Quantitative Sorbonne Université

Sanofi Campus Gentilly

4 Dec 2018



## All models are wrong, but some are useful.

[George E.P. Box, 1976]





rapidly accumulating...



#### Data..

## **Sequence data** are rapidly accumulating...



HOME | SEARCH | BROWSE | FTP | HELP | ABOUT

#### Pfam 30.0 (June 2016, 16306 entries)

The Pfam database is a large collection of protein families, each represented by **multiple sequence** alignments and hidden Markov models (HMMs). <u>More...</u>

QUICK LINKS	YOU CAN FIND DATA IN PFAM IN VARIOUS WAYS
SEQUENCE SEARCH	Analyze your protein sequence for Pfam matches
VIEW A PFAM ENTRY	View Pfam annotation and alignments
<b>VIEW A CLAN</b>	See groups of related entries
VIEW A SEQUENCE	Look at the domain organisation of a protein sequence
<b>VIEW A STRUCTURE</b>	Find the domains on a PDB structure
KEYWORD SEARCH	Query Pfam by keywords
JUMP TO	enter any accession or ID Go Example
	Enter any type of accession or ID to jump to the page for a Dfam entr

Enter any type of accession or ID to jump to the page for a Pfam entry or clan, UniProt sequence, PDB structure, etc.



- ...and organized into **protein families**:
- common evolutionary origin (homologs)
- conserved biological structure / function
- diverged sequences (20-30% seq ID)
- available as multiple-sequence alignments

#### opportunity for data-driven approaches

## Looking for information in :

-LNQFADDLAHELRTPVNILLGKNQVMLS-QERSAEEYQQALVDNIEELEGLSRLTENILFLARAEH-ALGELTAGIAHEINNPTAVILGNTELIRFLGADASRV-EEEIDAILLQIERIRNITRSLLQYSRQG--SORQFVTNASHELKTPIAIISANTEVLEI----TMGK-NOWTETILKQVKRLSGLVNDMVALAKLEE----AFVSNASHELRTPVTSIKGFAETIKG-MSAEEEAKDDFLDIIYKESLRLEHIVEHLLTLSKA0---VGQLTGGIAHDFNNMLTGVIGSLDLIKLS----GRLVERFMDAALISAQRAASLTDRLLAFSRRQS----RMTH0VSHEVGNMIGIITGSLGLLERETGFNDR0-KRHIARIRKAADRGRSLASSMLTIGS----ALGEMLDHIAHQWKQPINSISLIAQDMADYGELTDGDVQTTIDKIMSLLEHMSQTVDVFRGFYR-----VGRLAGGVAHDFNNLLSVINGYCEMLAA-0VSDRP0ALREVSEIHRAGLRAAGLTR0LLAFGRR0--SLGELAAGVAHEINNPNAVILLNVDLVKKWSEMSEEL-PLLLTEMEEGAGRIKRIVDDLKDFARGD---MGEFAAYIAHEINOPLSAIMTNANAGTRNEPSNIPEAKEALARIIRDSDRAAEIIRMVRSFLKRO-----GQLAGGIAHDFNNILQIISGNTQILQYQTNPDPP----QLLEILKAVERGTALTRSMLAFSRKQT----GQLTGGIAHDFNNLLQVILGNLEFVRAKLDGDAK-LQTRIERAAWAAQRGATLTGQLLAFARKQ---AKTDFLSNMSHEIRTPLNAILGFIQVLKD-AEMKPKD-REYLELMDESSKNLLSLVNDIIEIDLIESG ---GREVLHLVHDLKTPLATIEGLVSLMET-RWPDPKM-QEYCQTIYGSITSMSKMVSEILY-----RARLLADVAHELRTPVATLTGYLEAVEDVRPLDAST----IAVLRDQAVRLTRLAQDLADVTHAEGG SMKRMLTNMSHDLKTPLTVILGYIETI0SDPNMPDEERERLLGKLR0KTNELI0MINSFFDLAKLES-AKSEFLANMSHELRTPLNAIIGFSEMIQAFGPLGSDRYEEYINDIHTSGNFLLNVINDILDMSKIEAG -MQRFIADATHQLRTPLAAIDAEVELLTD-QTRDPKA----LDKLRGRIADLARLASQLLDHAM-----RKKAVHTIT<mark>H</mark>ELRTPLTAITG<mark>Y</mark>AGLIRK-EQCEDKS-GQYIQNILQSSDRMRDMLNTLLDFFRLDNG -REEFMNMTSHELMNPLSAAVOAAHTMISLHDDNSKSNIEIAKIILACGEHOQKLVEDARMMSKLD---KSRYVVGLSHELRSPLNAISGYA0LLEQDTSLAPKP-RD0VRVVRRSADHLSGLIDGILDISKIEAG ----AFSYMRHAINNPLSGMLYSRKALKN-TDLNEE0-MR0IHVSDNCHH0LNKILADL-----QENFIDMTSHEMRNPLSAILQCSDEITST----LCLEAANTIALCASHQKRIVDDILTFSKLDS-SORTLTNAIAHDLROPLYRIRFALEMFND-SLLSIEOROOYROSIENSLRDLDHLINOSLOLSRYT-----KLLLLSLSHDIKTPLSAIKLNAKALSRLYKDAEKQ-REAAEHINARADEIENFVSRITKASSE------HAFIADAAHELRTPLTALKLQLQLTER---ATSDVREVGFVKLNERLDRSIHLVKQLLTLARSES--QKNFISNASHELNTPLTSIIVTADLALS-KQRTDEEYRTALSRIMDAAGHLE--RGALLTSISHDLRTPLASILGATSSLESGEELDENARKELLSTIHDEADRLNRFVANLLDMTRLEAG -KSEFLANMSHELRTPLNGVIGFTRLTLK-TELTPTQ-RDHLNTIERSANNLLAIINDVLDFSKLEAG AKSEFLANMSHDIRTPMNAITGMTAIATA-HIDDPK0VKNCLRKIALSSRHLLGLINDVLDMSKIESG –LSQFSADLAHDFRTPLANLIGQTEVTLA–HPRSAEEYRAVLESSLEEYARLSRMIEDMLFLARADH– SKSMFLATVSHELRTPLYGIIGNLDLLQT-KELPKGV-DRLVTAMNNSSSLLLKIISDILDFSKIES-AKTAFLATLSHEIRTPMNGVLGTA0ILLK-TPLSTE0-EKHLKSLYDSGDHMMTLLNEILDFSKIE0G SKKQLIDGIAHELRTPLVRLRYRLEMSEN---LTPPE---SQALNRDIGQLEALIEELLTYARLDR--KTQFFINTAHDIRTPLTLIKAPLEELLEEETLTDNG-ITRTNIALRNVEVLLRLVSNLINFERT------VFIDNMTHEMKTPLTSIIGFSDLLRS-ARLDDETVHDYAESIYKEGKYLKSISSKLMDL------

#### multiple-sequence alignment

- 50-500 positions sequence length
- 10<sup>3</sup>-10<sup>5</sup> homologous sequences

## Inference of statistical sequence models

From data to statistical models

$$P(\overline{A}) = P(A_1, ..., A_L)$$

$$Data - MSA$$

$$\{\overline{A}^{\mu}\}_{\mu=1...M}$$
...

#### Attention: Data insufficient to do this without modeling

## Inference of statistical sequence models

From data over observables to statistical models

$$P(\overline{A}) = \frac{1}{Z} \exp\left\{\sum_{a} \lambda_{a} \mathcal{O}^{a}(\overline{A})\right\}$$

$$maximum \\ entropy \\ model$$

$$\langle \mathcal{O}^{a}(\overline{A}) \rangle_{P} = \frac{1}{M} \sum_{\mu} \mathcal{O}^{a}(\overline{A}^{\mu})$$

$$model average over data$$

#### Attention:

- model depends on data only via sample-averaged observables
- selection of observables requires prior biological knowledge

## **Conservation in proteins**





## All models are wrong, but some are useful.

[George E.P. Box, 1976]

#### **Profile models**



- identify conserved positions as functionally or structurally important positions
- are used to identify homologous sequences (automatic annotation)
- are used to align sequences

belong to the most successful statistical models in bioinformatics

## All models are wrong, but some are useful.

[George E.P. Box, 1976]

#### **Profile models**



- identify conserved positions as functionally or structurally important positions
- are used to identify homologous sequences (automatic annotation)
- are used to align sequences

belong to the most successful statistical models in bioinformatics

- are intrinsically unable to capture relations between positions
  - residue-residue contacts
  - protein-protein interaction

miss important information contained in sequence alignments

## Conservation and coevolution in proteins



## Conservation and coevolution in proteins



**Direct Coupling Analysis (DCA)** – pairwise residue-residue couplings

$$P(a_1, ..., a_L) \sim \exp\left\{ \underbrace{\sum_{i < j} J_{ij}(a_i, a_j)}_{i < j} + \sum_i h_i(a_i) \right\}$$

[Weigt et al, PNAS '09] [Morcos et al, PNAS '11]

## Direct coupling analysis (DCA)

- Boltzmann-machine learning:
  - start with initialised parameters (fields/couplings)
  - calculate

$$P_{ij}(A_i, A_j) = \sum_{\{A_k | k \neq i, j\}} P(A_1, ..., A_L)$$

• update parameters to fit marginals

$$\Delta J_{ij}(a,b) = \varepsilon [f_{ij}(a,b) - P_{ij}(a,b)]$$

- iterate until sufficiently precise fitting
- exact calculation requires exponential time ~ 21<sup>L</sup>
   approximations (MCMC) needed for computational efficiency

## Direct coupling analysis (DCA)

#### pseudo-likelihood maximisation

- replace ensemble average by sample average over sequence data

$$P_{1}(A_{1}) = \sum_{\{A_{i}|i>1\}} P(A_{1},...,A_{L})$$
  
=  $\sum_{\{A_{i}|i>1\}} P(A_{1}|A_{2},...,A_{L})P(A_{2},...,A_{L})$   
 $\simeq \frac{1}{M} \sum_{m=1}^{M} P(A_{1}|A_{2}^{m},...,A_{L}^{m})$   
biological sequence data

[Balakrishnan et al., Proteins 'I I] [Ekeberg, Lövkvist, Lan, MW, Aurell PRE 'I 3]

## Are pairwise DCA couplings useful?

DCA models are graphical statistical models:

- defined on a **network** of strongly coupled residues
- provide a **probability** to each sequence (sequence landscape)

What is the **biological information** contained in such models ?

## Strong couplings predict residue contacts



- works across numerous protein families
- accurate prediction for >1000 diverged sequences
- guides 3D protein structure prediction

[Marks et al., PLoS ONE '11] [Sulkowska, Morcos, <u>MW</u>, Hwa, Onuchic, PNAS '12] [Hopf et al., Cell '12] [Nugent, Jones, PNAS '12] [Ovchinnikov et al., eLife '15] [Ovchinnikov et al., Science '17]

## Prediction of inter-protein residue coevolution



## Are pairwise DCA couplings useful?

DCA models are graphical statistical models:

- defined on a **network** of strongly coupled residues
- provide a **probability** to each sequence (sequence landscape)

What is the **biological information** contained in such models ?

## Measuring mutational effects in proteins

# Capturing the mutational landscape of the beta-lactamase TEM-1 PNAS 110 (2013) 13067

Hervé Jacquier<sup>a,b,c,1</sup>, André Birgy<sup>a,b</sup>, Hervé Le Nagard<sup>a,b,d,e</sup>, Yves Mechulam<sup>f</sup>, Emmanuelle Schmitt<sup>f</sup>, Jérémy Glodt<sup>a,b</sup>, Beatrice Bercot<sup>c,g</sup>, Emmanuelle Petit<sup>h</sup>, Julie Poulain<sup>h</sup>, Guilène Barnaud<sup>i</sup>, Pierre-Alexis Gros<sup>a,b,j</sup>, and Olivier Tenaillon<sup>a,b,1</sup>



## Deep mutational scanning of proteins

#### **TEM-I** protein

causes antibiotic resistance

#### generated ~10<sup>4</sup> random mutants

- 1,700 without mutation
- 990 distinct single AA changes

#### measured resistance to amoxicillin

 minimum inhibitory concentration as proxy for fitness

## Landscape inference by Direct-Coupling Analysis

Beta-lactamase2 family (PFI3354)



Statistical landscape inference (DCA)

 $\sim \exp\left\{\sum_{i,j=1}^{L} e_{ij}(A_i, A_j) + \sum_{i=1}^{L} h_i(A_i)\right\}$ 

 $P(A_1, ..., A_L)$ 



[Figliuzzi, Jacquier, Schug, Tenaillon, MW, Mol Biol Evol '16]

## Landscape inference by Direct-Coupling Analysis

Beta-lactamase2 family (PFI3354)



[Figliuzzi, Jacquier, Schug, Tenaillon, MW, Mol Biol Evol '16]

## Predicting mutational effects in proteins



[Figliuzzi, Jacquier, Schug, Tenaillon, MW, Mol Biol Evol '16]





DMS: mutational data

[Barrat-Charlaix, Figliuzzi, MW, Sci Rep '16]

Statistical model

$$P(\overline{A}|\mathbf{J},\mathbf{h}) = \frac{1}{Z} \exp\left\{\sum_{a < b} J_{ij}(A_i, A_j) + \sum_a h_i(A_i)\right\} = \frac{1}{Z} \exp\left\{-\mathcal{H}(\overline{A})\right\}$$

Probability of MSA

$$P(\{\overline{A}^{\mu}\}_{\mu}|\mathbf{J},\mathbf{h}) = \exp\left\{-\sum_{\mu}\mathcal{H}(\overline{A}^{\mu}) - M\ln Z\right\}$$





DMS: mutational data

Statistical model

$$P(\overline{A}|\mathbf{J},\mathbf{h}) = \frac{1}{Z} \exp\left\{\sum_{a < b} J_{ij}(A_i, A_j) + \sum_a h_i(A_i)\right\} = \frac{1}{Z} \exp\left\{-\mathcal{H}(\overline{A})\right\}$$

Probability of MSA

$$P(\{\overline{A}^{\mu}\}_{\mu}|\mathbf{J},\mathbf{h}) = \exp\left\{-\sum_{\mu}\mathcal{H}(\overline{A}^{\mu}) - M\ln Z\right\}$$

Mutational data - Gaussian experimental noise

$$P(\{E^a\}_a | \{\overline{A}^a\}_a, \mathbf{J}, \mathbf{h})) = \frac{1}{(2\pi\Delta^2)^{P/2}} \exp\left\{-\frac{1}{2\Delta^2} \sum_a \left(E^a - \mathcal{H}(\overline{A}^a)\right)^2\right\}$$

Log-likelihood of model parameters

$$\mathcal{L}(\mathbf{J}, \mathbf{h}|data) = \log P(\{\overline{A}^{\mu}\}_{\mu}|\mathbf{J}, \mathbf{h}) + \log P(\{E^{a}\}_{a}|\{\overline{A}^{a}\}_{a}, \mathbf{J}, \mathbf{h})$$

#### Integrating heterogenous data in landscape inference 0.5 solvent accessibility 0.4 force fields 0.3 $\mathbb{R}^2$ Popmusic Blosum62 Imut+ 0.2 profile model mode PolyPhen2 MUpro 0.1 mut **∀** U SIFT 0 evolution based structural-stability based



[Barrat-Charlaix, Figliuzzi, MW, Sci Rep '16]

## Are pairwise DCA couplings useful?

DCA models are graphical statistical models:

- defined on a **network** of strongly coupled residues
- provide a **probability** to each sequence (sequence landscape)

What is the **biological information** contained in such models ?

## Are pairwise couplings sufficient?

Idea: scramble multiple-sequence alignments of homologs to conserve

- global amino-acid frequencies (no site specificity)
- site independent conservation (protein profile)  $\rightarrow$  only local fields, no couplings
- pairwise amino-acid co-occurrences  $\rightarrow$  pairwise residue-residue couplings



## Predicting folding sequences of the WW domain



## Data-driven statistical design

.

. . .

From data over observables and models to data

$$P(\overline{S}) \sim \exp\left\{\sum_{a} \lambda_{a} \mathcal{O}^{a}(\overline{S})\right\} \longrightarrow \qquad \{\overline{X}^{\nu}\}_{\nu=1,...,N}$$

$$(\mathcal{O}_{a}(\overline{S}))_{P} \simeq \frac{1}{M} \sum_{\mu} \mathcal{O}_{a}(\overline{S}^{\mu}) \longleftarrow \qquad \{\overline{S}^{\mu}\}_{\mu=1,...,M}$$

## Data-driven statistical design

From data over observables and models to data



. . .

## DCA reaches very precise fitting response regulator (PF00072)



tested for many protein families

[Figliuzzi, Barrat-Charlaix, MW, Mol. Biol. Evol. 2018]

## DCA reproduces non-fitted quantities

response regulator (PF00072)



➡ tested for many protein families

[Figliuzzi, Barrat-Charlaix, MW, Mol. Biol. Evol. 2018]

## Coevolution-guided protein design



experiments by B. Russ, R. Ranganathan

### ...all models are wrong, but some are useful. [George E.P. Box, 1976]

## Biological sequence evolution ≠ MCMC sampling from Potts models

### ...all models are wrong, but some are useful.

[George E.P. Box, 1976]

#### However DCA allows for

- protein structure prediction strong couplings accurately predict contacts
- mutational effect prediction statistical model as sequence landscape
- generative statistical modeling pairwise couplings seem necessary and sufficient to capture biological sequence diversity

## ...all models are wrong, but some are useful.

#### [George E.P. Box, 1976]

#### However DCA allows for

- protein structure prediction strong couplings accurately predict contacts
- mutational effect prediction statistical model as sequence landscape
- generative statistical modeling pairwise couplings seem necessary and sufficient to capture biological sequence diversity

#### A few current limitations

- many parameters vs. limited data strong regularization against overfitting
  - parsimonous modeling / dimensional reduction ?
- correlated data DCA fits functional and phylogenetic correlations
   inference from non-i.i.d. samples ?
- better models may be even more useful...
  - systematic selection of statistically relevant observables ?
- uses multiple-sequence alignment based on profile models
  - more accurate methods using couplings for alignment ?
- sequences modeled as strings over abstract 21-letter alphabet
  - prior biological knowledge / integrative modeling ?

## Thanks to:

The group in Paris:

Juliana Bernardes Pierre Barrat-Charlaix Giancarlo Croce Kai Shimagaki Edwin Rodriguez Nika Abdollahi Anna-Paola Muntoni Maureen Muscat

Francesco Oteri

Alumni:

Maria Virginia Ruiz Cuevas Eleonora de Leonardis Guido Uguzzoni Alice Coucke Matteo Figliuzzi Christoph Feinauer

Funding:







Terry Hwa (UC San Diego) Hendrik Szurmant (Western U LA) Alexander Schug (KIT Karlsruhe) Jose Onuchic (Rice U, Austin) Faruck Morcos (UT Dallas) Angel E. Dago (Scripps La Jolla) Joanna Sulkowska (U Warsaw) Erik Aurell (KTH Stockholm) Andrea Pagnani (Politecnico Torino) Thomas Gueudré (IIGM Torino) Carlo Baldassi (U Bocconi Milano) Rémi Monasson (ENS Paris) Simona Cocco (ENS Paris) Olivier Tenaillon (Inserm Paris) Bill Russ (UTSW Dallas) Rama Ranganathan (U Chicago) Anne-Florence Bitbol (Sorbonne U Paris) Francesco Zamponi (ENS Paris)





Horizon 2020 European Union funding for Research & Innovation